



## Short report

## Radical scavenging, antioxidant and cytotoxic activity of Brazilian *Caatinga* plants

Juceni P. David<sup>a,\*</sup>, Marilena Meira<sup>b</sup>, Jorge M. David<sup>b</sup>, Hugo N. Brandão<sup>b</sup>,  
Alexsandro Branco<sup>a</sup>, M. de Fátima Agra<sup>c</sup>, M. Regina V. Barbosa<sup>d</sup>,  
Luciano P. de Queiroz<sup>e</sup>, Ana M. Giulietti<sup>e</sup>

<sup>a</sup> Faculdade de Farmácia, Universidade Federal da Bahia, 40170-290 Salvador-BA, Brazil

<sup>b</sup> Instituto de Química, Universidade Federal da Bahia, 40170-290 Salvador-BA, Brazil

<sup>c</sup> Laboratório de Tecnologia Farmacêutica, Universidade Federal da Paraíba, 58051-970 João Pessoa-PB, Brazil

<sup>d</sup> Departamento de Sistemática e Ecologia, Universidade Federal da Paraíba, 58051-970 João Pessoa-PB, Brazil

<sup>e</sup> Departamento de Ciências Biológicas, Universidade Estadual de Feira de Santana, 44031-460, Feira de Santana-BA, Brazil

Received 8 November 2005; accepted 14 November 2006

Available online 6 February 2007

### Abstract

Extracts of 32 plants from the Brazilian northeastern semi-arid region called *Caatinga* were evaluated through DPPH radical scavenging assay, β-carotene bleaching, and brine shrimp lethality tests (BST). Among the extracts studied *Byrsonima cf. gardneriana*, *Mascagnia coriacea*, *Cordia globosa*, *Diodia apiculata* and *Hypenia salzmannii* showed the highest activities in DPPH radical scavenging test. In the β-carotene bleaching test the highest activities were observed for *Passiflora cincinnata*, *Chamaecrista repens*, *B. cf. gardneriana*, *Rollinia leptopetala*, *Serjania glabrata*, *Diospyros gaultherifolia*, *C. globosa*, *Mimosa ophtalmocentra*, *M. coriacea* and *Lippia cf. microphylla*. In contrast, *R. leptopetala*, *Zornia cf. brasiliensis* and *Leonotis nepetifolia* were the most active species in the BST. © 2007 Elsevier B.V. All rights reserved.

**Keywords:** Antioxidant activity; Brine shrimp test; Brazilian medicinal plants

### 1. Plants

Plant species listed in Table 1 were collected in the region called *Caatinga* in northeastern Brazil in March 2003. A voucher of each species is deposited in the Herbarium Lauro Pires Xavier in the Universidade Federal da Paraíba and Universidade Estadual de Feira de Santana's Herbarium.

### 2. Uses in traditional medicine

The plants studied are employed as medicine by the local population in the Brazilian northeastern region. They are used as laxative, sedative, for dermatitis, anti-inflammatory, colds, asthma, eczema, influenza, anti-

\* Corresponding author. Tel.: +55 7132359350; fax: +55 7132355166.

E-mail address: [juceni@ufba.br](mailto:juceni@ufba.br) (J.P. David).

Table 1

Radical scavenging, antioxidant and cytotoxic activity of the methanol extracts of *Caatinga* plants

Plants	Part used	Yield extract (%)	Radical scavenging activity (DPPH assay)		Antioxidant activity ( $\beta$ -carotene bleaching test)		BST (brine shrimp lethality test)	
			IC <sub>50</sub> in mg/ml	95% Confidence interval	% AA at 10 mg/ml	95% Confidence interval	LC <sub>50</sub> in $\mu\text{g}/\text{ml}$	95% Confidence interval
<i>Acnistus arborescens</i> L., Schltdl. (Solanaceae), (Agra 5790)	Aerial parts	1.4	16.2	14.5–18.4	34.8	25.1–44.5	141.4	129.0–155.3
<i>Allamanda blanchetii</i> A. DC. (Apocynaceae), (Agra 5673)	Aerial parts	1.7	3.7	3.3–4.3	47.1	27.4–66.7	373.1	227.0–601.6
<i>Blainvillea rhomboidea</i> Cass. (Asteraceae), (Agra 5888)	Aerial parts	4.0	10.7	9.6–12.0	36.2	30.8–41.6	426.2	394.8–462.5
<i>Byrsinima gardneriana</i> A. Juss. (Malpighiaceae), (Agra 947)	Aerial parts	10.6	0.3	0.09–0.7	63.2	56.7–69.7	243.9	220.9–269.9
<i>Cardiospermum corindum</i> L. (Sapindaceae), (Agra 5858)	Aerial parts	7.0	2.3	2.0–2.6	41.4	33.4–49.5	142.9	108.9–188.3
<i>Chamaecrista repens</i> (Vog.) H.S. Irwin et Barneby var. <i>multijuga</i> (Benth.) H.S. Irwin et Barneby (Caesalpiniaceae), (Agra 5924)	Aerial parts	6.1	2.0	1.3–3.1	68.3	61.5–75.0	269.3	256.0–282.9
<i>Cnidoscolus phyllacanthus</i> Pax et Hoff. (Euphorbiaceae), (Agra 5997)	Aerial parts	1.5	13.2	12.2–14.3	39.3	21.8–56.8	89.7	63.2–127.3
<i>Cordia globosa</i> (Jacq.) Kunth. (Boraginaceae), (Agra 4969)	Aerial parts	8.5	1.03	0.96–1.11	55.0	45.8–64.2	201.0	112.6–361.0
<i>Cordia multispicata</i> Cham. (Boraginaceae), (Agra 5741)	Aerial parts	2.5	2.2	1.8–2.6	42.7	35.3–50.0	227.5	145.8–354.4
<i>Crescentia cujete</i> L. (Bignoniaceae), (Agra 5838)	Aerial parts	3.5	10.3	7.1–16.6	38.3	24.5–52.1	723.3	471.0–1109.2
<i>Croton moritibensis</i> Baill. (Euphorbiaceae)	Aerial parts	9.6	5.5	4.8–6.4	45.4	33.0–57.9	31.0	15.7–40.8
<i>Diodia apiculata</i> (R. et S.) K. Schum. (Rubiaceae), (Agra 5874)	Aerial parts	4.9	1.3	1.2–1.6	20.1	16.5–23.8	459.1	310.0–678.7
<i>Diospyros gaultheriifolia</i> Mart. et Miq. (Ebenaceae), (Agra 5626)	Aerial parts	4.6	5.5	4.1–7.6	56.1	52.4–59.9	883.0	767.1–1015.4
<i>Herissantia crispa</i> L. (Brizicky) (Malvaceae), (Agra 5868)	Aerial parts	3.1	3.9	3.5–4.4	45.6	28.8–62.3	281.2	155.2–515.6
<i>Hypenia salzmannii</i> (Benth.) Harley (Lamiaceae), (Agra 5926)	Leaves and stems	7.0	1.5	1.4–1.7	25.8	22.0–29.6	159.1	131.7–192.9
<i>Ipomea hederifolia</i> L. (Convolvulaceae), (Agra 5838)	Leaves and stems	16.0	2.7	2.5–2.9	33.7	29.3–38.2	177.9	114.0–278.9
<i>Ipomea martii</i> Meissn. (Convolvulaceae), (Barbosa 2249)	Aerial parts	4.9	4.4	3.9–4.9	34.5	20.4–48.6	125.6	82.1–192.4
<i>Jacquemontia densiflora</i> (Convolvulaceae), (Agra 5923)	Aerial parts	6.4	4.1	3.8–4.5	44.6	40.8–48.5	193.8	76.2–491.1
<i>Lantana canescens</i> Kunth (Verbenaceae), (Agra 5859)	Aerial parts	3.4	3.1	2.5–3.9	36.9	33.3–40.4	127.4	52.2–311.0
<i>Leonotis nepetifolia</i> (L.) R. Br. (Lamiaceae), (Agra 5621)	Aerial parts	3.0	6.5	5.7–7.4	47.9	33.9–62.0	19.8	13.5–28.9
<i>Lippia cf. microphylla</i> Cham. (Verbenaceae), (Agra 5693)	Leaves and stems	5.6	3.2	2.9–3.4	50.1	39.6–60.5	73.7	63.9–84.9
<i>Mascagnia coriacea</i> Griseb. (Malpighiaceae), (Agra 5836)	Aerial parts	6.0	0.3	0.31–0.37	50.2	33.7–66.7	423.7	215.2–838.7
<i>Mimosa ophtalmocentra</i> Mart. ex Benth. (Mimosaceae), (Agra 5933)	Aerial parts	12.3	6.2	5.9–6.5	52.2	34.8–69.7	185.1	103.5–325.3
<i>Nicandra physalodes</i> (L.) Gardn. (Solanaceae), (Agra 5891)	Leaves and stems	3.9	4.2	3.4–5.4	46.7	43.9–49.4	48.2	30.4–75.1
<i>Passiflora cincinnata</i> Mast. (Passifloraceae), (Agra 5873)	Aerial parts	4.5	22.3	18.8–27.3	70.1	57.6–82.6	266.9	193.2–369.4
<i>Pilocarpus spicatus</i> A. St.-Hill. (Rutaceae), (Agra 5788)	Leaves and stems	11.7	3.6	2.9–4.4	38.6	34.9–42.2	95.2	61.5–148.4
<i>Rollinia leptopetala</i> R. E. Fr. (Annonaceae), (Agra 5700)	Leaves and stems	5.5	3.5	3.1–4.0	57.3	44.9–69.7	0.6	0.2–1.8
<i>Serjania glabrata</i> Kunth (Sapindaceae), (Barbosa 2266)	Aerial parts	1.1	2.8	2.6–3.1	57.1	48.8–65.5	248.0	197.4–313.9
<i>Sidastrum paniculatum</i> (L.) Fryxell (Malvaceae), (Agra 586)	Aerial parts	2.3	5.3	5.0–5.7	39.2	32.5–46.0	317.2	251.6–400.0
<i>Tocoyena formosa</i> (Cham. et Schltdl.) (Rubiaceae), (Agra 5734)	Leaves and stems	3.1	13.2	11.9–14.7	43.7	34.8–52.7	332.0	267.6–413.0
<i>Vernonia chalybaea</i> Mart. ex D.C. (Asteraceae), (Agra 5835)	Aerial parts	5.8	6.0	5.7–6.3	17.7	8.6–26.8	377.8	228.4–662.5
<i>Zornia cf. brasiliensis</i> Vog. (Fabaceae), (Barbosa 2235)	Aerial parts	2.1	25.1	19.0–35.8	46.6	43.1–50.2	7.2	4.5–11.4

hemorrhoid, diuretic, anti-rheumatism, for menstrual disorders, anti-hypertensive, digestive, hair tonic, and kidney disease [1,2].

### 3. Previously isolated classes of constituents

*A. arborescens* [3], *C. phyllacanthus* [4,5], *Cordia globosa* [6], *T. formosa* [7], *P. spicatus* [8,9] and *C. cujete* [10] were previously examined for their biologically activities and chemical constituents. In these studies were isolated cytotoxic withanolides and favelines, antifungal iridoids, coumarin with gGAPDH inhibitory activity and cytotoxic naphtoquinones, respectively. *B. rhomboidea* was found to contain sesquiterpene lactones, while *Rollinia leptopetala* contains dammarane derivatives and alkaloids [11,12].

### 4. Tested material

Methanol extracts were prepared at room temperature.

### 5. Studied activity

Radical scavenging activities of plant extracts were determined through spectrophotometry using 1,1-diphenyl-2-picrylhydrazyl (DPPH) scavenging radical assay [13]. The antioxidant activity of MeOH extracts was evaluated with the  $\beta$ -carotene bleaching test in a linolenic acid suspension as previously described with slight modifications [14]. Brine shrimp lethality test was performed according to Serrano et al. [15] with minor modifications [16].

### 6. Statistical analysis

All assays were developed in triplicate and the test results were analyzed using the two-tailed Student's *t*-test at a significance level of  $P < 0.05$  and DPPH IC<sub>50</sub> values with 95% confidence intervals were determined using the regression method with the Analyse-it software (Table 1). BST LC<sub>50</sub> values with a 95% confidence interval were determined using the probit analysis method of Stats Direct statistical software (Table 1). When required the results were found by extrapolation of the straight line.

### 7. Results and discussion

Among the extracts studied (Table 1) the highest scavenging activities (lowest IC<sub>50</sub>) were observed for *Byrsonima gardneriana* (0.3 mg/ml), *Mascagnia coriacea* (0.3 mg/ml), *C. globosa* (1.03 mg/ml), *Diodia apiculata* (1.3 mg/ml) and *Hypenia salzmannii* (1.5 mg/ml) when compared with the butylated hydroxyanisole (BHA) [IC<sub>50</sub> (34.1 ± 0.6) × 10<sup>-3</sup> mg/ml].

The activity was evaluated measuring the protection against oxidation in  $\beta$ -carotene bleaching test at a concentration of 10.0 mg/ml. The highest activities were observed for *Passiflora cincinnata* (70.1), *Chamaecrista repens* (68.3), *B. gardneriana* (63.2), *R. leptopetala* (57.3), *Serjania glabrata* (57.1), *Diospyros gaultheriifolia* (56.1), *C. globosa* (55.0), *Mimosa ophtalmocentra* (52.2), *M. coriacea* (50.2) and *Lippia microphylla* (50.1) (Table 1). The butylated hydroxytoluene (BHT) (1.0 mg/ml) presented AA of 84.3 ± 13.9.

The extracts with LC<sub>50</sub> values higher than 200 mg/l in the brine shrimp test can be considered inactive [17]. In accordance with this criterion, about 50% of the species presented positive results (LC<sub>50</sub> < 200 mg/l) (Table 1). However, other authors [15] consider this value as low. The most active species in BST were *R. leptopetala* (0.6 µg/ml), *Zornia cf. brasiliensis* (7.2 µg/ml) and *L. nepetifolia* (19.8 µg/ml).

### 8. Conclusions

No extracts were found to exhibit comparable radical scavenging or antioxidant activities with commercial antioxidants. Nevertheless, *P. cincinnata*, *C. repens*, *B. gardneriana*, *S. glabrata*, *D. gaultheriifolia*, *C. globosa* and *M. coriacea* showed AA and low toxicity in the BST.

## Acknowledgements

The authors thank CAPES, CNPq and the Instituto do Milênio do Semi-Árido (IMSEAR) for fellowships and financial support.

## References

- [1] Corrêa MP. Dicionário das Plantas Úteis do Brasil e das Exóticas Cultivadas, vol. I–VI. Rio de Janeiro: Ed. Imprensa Nacional; 1984.
- [2] Agra MDF. Plantas da Medicina Popular dos Cariris Velhos. João Pessoa, Paraíba: Ed. União; 1996. p. 125.
- [3] Minguzzi S, Barata LES, Shin YG, Jonas PF, Chai HB, Park EJ, et al. Phytochemistry 2002;59:635.
- [4] Endo Y, Ohta T, Nozoe S. Tetrahedron Lett 1992;33:353.
- [5] Ohta T, Endo Y, Kikuchi R, Kabuto C, Harada N, Nozoe S. Tetrahedron 1994;50:5659.
- [6] Silva SAS, Rodrigues MSL, Agra M de F, Cunha EL, Barbosa-Filho JM, Silva MS. Biochem Syst Ecol 2004;32:359.
- [7] Bolzani VS, Izumisawa CM, Young MCM, Trevisan LMV, Kingston DGI, Gunatilaka AL. Phytochemistry 1997;46:305.
- [8] Pavão F, Castilho MS, Pupo MT, Dias RLA, Correa AG, Fernandes JB. FEBS Lett 2002;520:13.
- [9] Andrade-Neto M, Silveira ER, Braz-Filho R, Gambardela MTP, Santos RHA. Phytochemistry 1994;35:739.
- [10] Kaneko T, Ohtani K, Kasai R, Yamasaki K, Duct NM. Phytochemistry 1998;47:259.
- [11] Spring O, Zipper R, Vogler B, Lopes JLC, Vichnewski W, Dias DA, et al. Phytochemistry 1999;52:79.
- [12] Sette IMF, Cunha EVL, Barbosa-Filho JM, Agra M de F, Silva MS. Biochem Syst Ecol 2000;28:393.
- [13] Hatano T, Kagawa H, Yasuhara T, Okuda T. Chem Pharm Bull 1988;36:2090.
- [14] Barreiros ALSB, David JP, Queiroz LP, David JM. Phytochemistry 2001;55:805.
- [15] Serrano C, Ortega T, Villar A. Phytother Res 1996;10:118.
- [16] David JP, Silva EF, Moura DL, Guedes MLS, Assunção RJ, David JM. Quim Nova 2001;24:730.
- [17] Anderson JE, Goetz CM, McLaughlin JL, Suffness MA. Phytochem Anal 1991;2:107.