

Estuarine ichthyofauna of the Paraguaçu River, Todos os Santos Bay, Bahia, Brazil

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Abstract: Estuaries are areas of recognized importance in the development and sanctuaries of fishes and also a great source of fishing resources. This article presents an inventory of fishes from the Paraguaçu River estuary, Todos os Santos Bay, Bahia, Brazil. 28 points comprising the saline sections that may exist in a tropical estuary were sampled quarterly from March 2009 to February 2010 with the aid of bottom gill nets, surface gill nets, seine net and casting net. A total of 4.097 individuals were captured, belonging to 7 species of Chondrichthyes and 117 species of Actinopterygii (83 genera and 49 families). Most of the geographical extent of the estuary was sampled by assessing large spatial variability in fish assemblage structure. The curve of species accumulation (rarefaction) had stabilization with ½ of the samples. The general structure of the assemblage is described and commented through evidences for a new spatial organization of the fish fauna due to dam effects upstream the estuary.

Keywords: inventory, estuarine fishes, spatial distribution, effects upstream, brazilian northeast.

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Resumo: Estuários são áreas de reconhecida importância no desenvolvimento e refúgio dos peixes e também grande fonte de recursos pesqueiros. Este artigo apresenta um inventário dos peixes do estuário do Rio Paraguaçu, Baía de Todos os Santos, Bahia, Brasil. 28 pontos constituindo as três seções salinas que pode haver em um estuário tropical foram amostradas trimestralmente de março de 2009 a fevereiro de 2010 com auxílio de redes de espera de fundo, de superfície, redes de arrasto manual e tarrafas. Um total de 4.097 indivíduos foram capturados, pertencendo a 7 espécies de Chondrichthyes e 117 espécies de Actinopterygii (83 gêneros e 49 famílias). A maioria da extensão territorial do estuário foi amostrada para acessar maior variabilidade espacial na estrutura da assembléia de peixes. A curva de acumulação de espécies (rarefação) apresentou estabilização com ½ das amostras. A estrutura geral da assembléia é descrita e comentada através de evidências para uma nova organização espacial da fauna de peixes devido a efeitos de barragem a montante do estuário.

Palavras-Chaves: inventário, peixes estuarinos, distribuição espacial, efeitos a montante, nordeste brasileiro.

Introduction

The Paraguaçu River, the main tributary of the Todos os Santos Bay (BTS), is one of the most important aquatic systems of the Bahia State. This system is of high value for wildlife conservation and provides the main source of protein and income (i.e. consumption and commercialization of fish and shellfish) for the local communities (Barros et al. 2008). In spite of the ecological and economic importance of Paraguaçu River estuary (127.9 km²), this river is located on the west side of Todos os Santos Bay and its origin is related to the drowning of a fluvial valley, during Middle-Holocene, controlled by the system of faults from the western portion of Recôncavo Bay, especially for the Maragojipe Fault (Carvalho 2000). The estuary is composed by three sectors: the lower course of the river (with 16 km long and average depth of 1 m), Iguape Bay (with 76.1 km² and an intertidal area of 53.7%) and by the Paraguaçu channel (with 18 km long and a average depth of 10 m), which establishes a connection between the Iguape Bay and Todos os Santos Bay, associated with the good state of conservation and the presence of a wide mangrove area (28 km²), favoring a high fishing productivity and the presence of a wide riverside community involved with artisanal fishing (Genz et al. 2008). The municipality of Maragojipe, alongside the Iguape Bay, has come to be the second major fish producer in the Bahia State (BAHIAPESCA 1998), where in 2000 the IBAMA created the Iguape Bay Marine Extractive Reserve (Decree no number of 14/08/2000). During the 80's, the Pedra do Cavalo Dam (PCD) was built about 28 km from de mouth of Iguape Bay, causing changes on the hydrological regime of the river. In 2005, a new change occurred due to the implantation of the Pedra do Cavalo Hydroelectric Power Plant (HPP), changing the fresh water flow to energy generation (Genz et al. 2008). Nowadays there is a project in licensing for deployment of a port system within the estuary where extensive mangrove zone and estuarine sediment will be removed, setting one more impact scenario for the fish fauna as well as all the marine biota.

Few studies with the clear objective of surveying the fish fauna of the estuaries on Northeastern Brazil have been carried (Eskinazi 1970, 1972, Oliveira 1972, 1974, 1976, Araújo et al. 2000). Most studies focus on ecological aspects of ichthyofauna from shallow areas (Alves & Soares Filho 1996, Castro 2001, Barletta et al. 2003, Carvalho Neta & Castro 2008, Paiva et al. 2008, Reis-Filho et al. 2010), fishes as accompanying fauna in shrimp fisheries (Barros & Jonsson 1967, Albuquerque 1994, Santos et al. 1998, Santos 2000) and few with deep drags targeted for capture of fishes (Barletta et al. 2005). Thus, studies that seek access the fish fauna evaluating the variability of habitats of the estuarine system are missing.

In the Paraguaçu River estuary there is only one study approaching communities of fish in the mouth (Reis-Filho, in press), and a work addressing expansion of geographical distribution (Santos et al. 2009). Studies of faunal surveys constitute the initial step for biological studies and management of an area, by providing basic information on the composition and structure of the investigated fauna (Casatti et al. 2001). However, the fact of the Todos os Santos Bay being the second largest semi-enclosed bay in Brazil and have no ichthyofaunal study in their estuaries (Andrade-Tubino et al. 2008), besides the fact that studies reported for Brazilian estuaries are credited only to the north and south region (Faunce & Serafy 2006) sets up such a worrying scenario. The urgent need to know the ichthyofauna of the place so as to develop conservation and management measures corroborates the lack of these studies in estuarine environments. This way, the estuarine ichthyofauna from Paraguaçu River, Todos os Santos Bay, Bahia was studied the main factors that influence the distribution and composition, by making a parallel with the effect of dam upstream the estuary.

Material and Methods

Fishes were collected at 28 points within the Paraguaçu River estuary with quarterly samples between March 2009 and February 2010 (Figure 1). Several types of collectors were used: for marginal environments and with depths up to 3 m, trawl manual (12 m × 2 m with 12 mm mesh), gillnets (100 m × 3 m with 15 mm mesh) and flue (casting net) (10 mm mesh). For environments with depth between 4 and 20 m, bottom gill nets (300 m × 3 m with 20 mm, 30 mm, 40 mm, 50 mm and 60 mm mesh) and surface gill nets (200 m × 4 m with 15 mm mesh) were used. In order to evaluate the ichthyofauna distribution in the estuary, this one was segmented into three sections (lower, middle and upper).

Specimens were transported in ice and stored frozen until identification with the most current literature (Figueiredo & Menezes 1978, 1980, 2000, Menezes & Figueiredo 1980, 1985, Carpenter 2002a, b, Marceniuk 2005; Moura & Lindeman 2007). Total length (mm) and weight (g) was recorded for each specimen. Vouchers were deposited at the Universidade Federal da Bahia (UFBA). The occurrence of constancy of species was calculated as function of the number of times each species took place in relation to the total collections done (Dajoz 1973). In order to identify the dominant species, we used the Relative Importance Index (IRI) as defined by Beaumond (1991), where N_i is the number of species individuals, W_i is the weight of the specimens of the species and C is the constancy of occurrence (Figure 2). The efficiency of the inventory was assessed by the method of species accumulation (rarefaction) at program PAST® (Paleontological Statistics, ver. 1.75). A multivariate analysis of type DCA (Detrended Correspondence Analysis) (ter Braak 1995) was applied to the data of species abundance dominant and constant, transformed into neperian logarithm to synthesize community variation along the spatial gradient. The salinity was measured at all points during the sampling campaigns with the aid of optical refractometer in order to define the different saline zones within the estuary.

The fish species were classified in five estuarine-use functional groups proposed by Elliott et al. (2007): 1) marine stragglers, species that spawn at sea and typically enter estuaries only in low numbers most frequently in the lower reaches and occur where salinities are c. 35; 2) marine migrants, species that spawn at sea and often enter estuaries in large numbers, particularly as juveniles. Some of these species are highly euryhaline and move throughout the full length of the estuary; 3) estuarine residents, estuarine species capable of completing their entire life cycle within the estuary environment; 4) estuarine migrants, estuarine species that have larval stages of their life cycles completed outside the estuary or are also represented by discrete marine or freshwater populations; and 5) semi-catadromous, species whose spawning run extends only to estuarine areas rather than the marine environment. The scientific nomenclature followed Nelson (1994), Eschmeyer (2006), Froese & Pauly (2006), Marceniuk & Menezes (2007), Marceniuk (2005) and Menezes et al. (2003). Classification according to habitat use was compared with species listed on the work of Barletta-Bergan (1999), Camargo (1999), Barletta et al. (2003, 2005, 2008), Fisher et al. (2004) and Moraes et al. (2009).

Results

A slight salt gradient was observed in the Paraguaçu estuary, with values above 35 near the mouth dropping to 27 in almost all the middle estuary (Iguape Bay). From this area the salinity has values increasingly smaller until it reaches 0 (zero), where the upper estuary starts. In this last sector, the salinity ranged from 0 to 5 as the movement of tides. The estuary of the Paraguaçu River during the sampling campaigns showed a behavior predominantly saline.

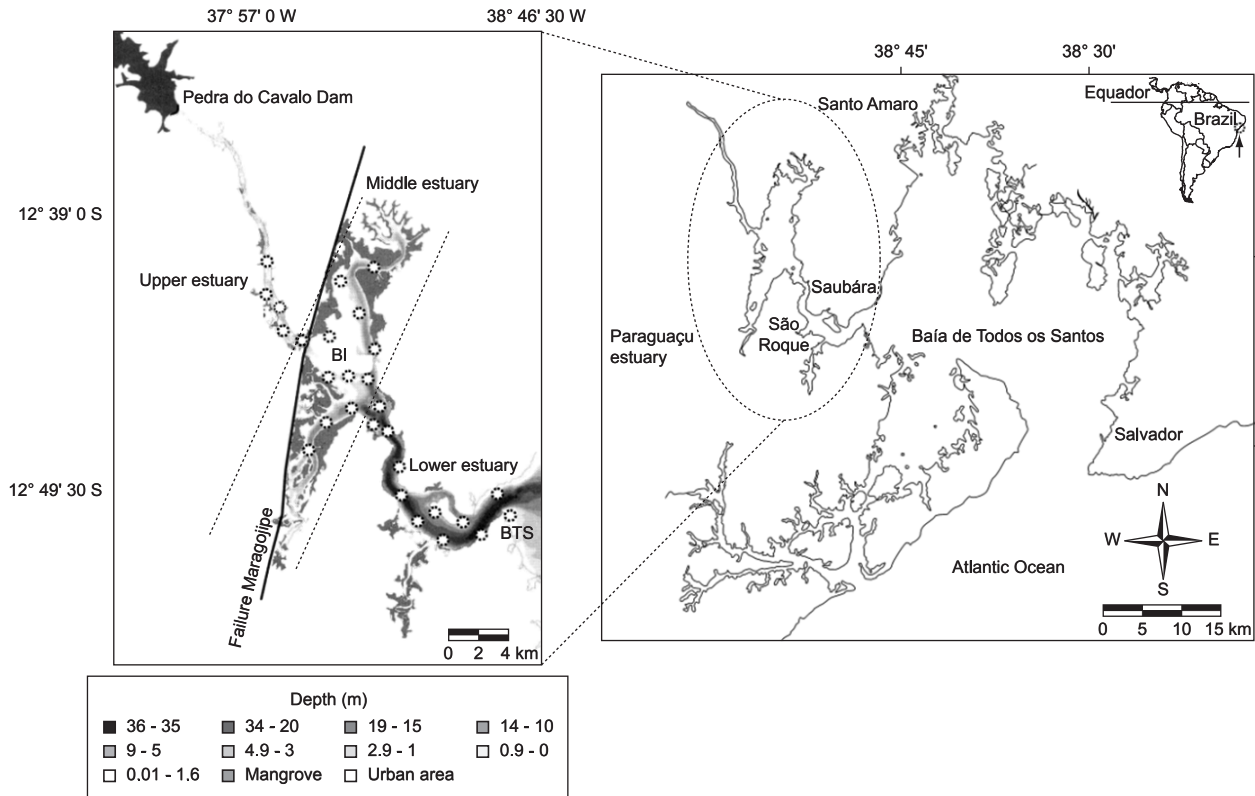


Figure 1. Location of Paraguaçu River estuary with sampling points. IB: Iguape Bay. (Adepted of Genz et al. 2008).

Figura 1. Localização do estuário do Rio Paraguaçu com pontos amostrados. IB: Baía de Iguape. (Adaptado de Genz et al. 2008).

$$IRI = \frac{(NiWiC)}{\sum(NiWiC)} \times 100$$

Figure 2. Formula used to calculate the Index of Relative Importance.

Figura 2. Fórmula usada para calcular o Índice de Importância Relativa.

The species accumulation curve, obtained by the technique of rarefaction of samples indicates that the samples with ½ the number of species seems to reach a plateau-linear with most species being accessed in these catches (Figure 3). A total of 224 samples were taken and 4,097 specimens were captured, belonging to 7 species of Chondrichthyes and 117 species of Actinopterygii (83 genera and 49 families). Most recorded species (62%) are definitely marine fishes, but a considerable number of estuarine species was also recorded (35%). Four Actinopterygii were identified only at the level of genus (*Anchoa* sp., *Paralichthys* sp., *Stellifer* sp. and *Mugil* sp.). Perciformes (IRI = 61.5%) was the most important order, followed by Clupeiformes (IRI = 18.2%), Pleuronectiformes (IRI = 11.2%) and Tetraodontiformes (7.9%). Sciaenidae e Carangidae were the most abundant families in number of species (12 each). Sciaenidae showed the most weight (39.8%). Sciaenidae and Gobiidae were the most abundant families in number of individuals (14.8% and 11.9%, respectively), followed to Gerreidae (9.4%), Engraulidae (8.2%), Clupeidae (5.5%) and Tetraodontidae (5.3%), together they accounted more than 55% of total abundance (Table 1). Among the nine most abundant and constant species, have been two of sciaenids, *Stellifer rastriifer* (Jordan, 1889) and *Cynoscion microlepidotus* (Cuvier, 1830) dominating the middle estuary, two of gerreids *Diapterus rhombeus* (Cuvier, 1829) and *Eucinostomus argenteus* Baird & Girard, 1855 abundant and frequent in the lower and middle estuary, one of Tetraodontidae, *Sphoeroides greeleyi* Gilbert, 1900 abundant

and constant in all three sectors and one Pristigasteridae, *Pellona harroweri* (Fowler, 1917) abundant and constant in the lower and middle estuary, and these were responsible for 52.5% of the expressed dominance (IRI).

It was possible to identify assemblages with distinct patterns of habitat use according to the zones of depths, behavior in the estuary and salt sectors. The dispersal of species along the axis 1 of the DCA, held in the Paraguaçu River estuary, indicates that the species exclusively caught in shallow areas have high abundances in all saline sectors of the estuary and the species caught in deep water are more abundant in the middle estuary (Figure 4a). Yet, the resident estuarine species show preference for the middle estuary with higher abundances recorded in this sector and reduced catches in the lower estuary. The marine migrants were found in all sectors and marine stragglers were from the mouth to the middle estuary with homogeneous densities among sectors (Figure 4b).

Discussion

The trend in stabilization of the species accumulation curve with about ½ of the samples corroborates with the information that in the estuarine environment the fish assemblage is dominated by few species (Day et al. 1989, Santos et al. 2002, Godefroid et al. 2004) and its basic structure may appear relatively stable or even predictable (Blaber & Blaber 1980, Whitfield 1999, Paterson & Whitfield 2000). One positive point for the stabilization of the accumulation curve was the use of various sampling techniques that have covered a great variety of habitats on the spatial gradient (e.g. shallow and deep zones, tidal channels and tidal flats). Among other aspects, the limited use of sample artifacts, especially towed passive network (gillnets) on the American continent, are responsible for poor knowledge about the habitat use by estuarine fish fauna (Faunce & Serafy 2006).

Table 1. Checklist of fish species captured at the Paraguaçu River estuary, Bahia, Brazil. N = Total Number of individuals; N (%) = Relative abundance; W (%) = Relative weight; C = Occurrence constancy; IRI = Importance relative index; HU = Habitat use; MS = Marine stragglers; ES = Estuarine residents; MM = Marine migrants; EM = Estuarine Migrants; and SC = Semi-catadroms. * = marine species found in the middle estuary.

Tabela 1. Lista das espécies de peixes capturadas no estuário do Rio Paraguaçu, Bahia, Brazil. N = Número total de indivíduos; N(%) = Abundância relativa; W(%) = Peso relativo; C = Constância de Ocorrência; IRI = Índice de importância relativa; HU = Uso do habitat; MS = Visitantes Marinhos; ES = Estuarinos residentes; MM = Migrantes Marinhos; EM = Migrantes Estuarinos; e SC = Semi-catádroms. * = espécies marinhas encontradas no estuário médio.

| Class, Order, Family and Species | N | N (%) | W (%) | C (%) | IRI | HU |
|--|-----|-------|-------|-------|-------|-----|
| Class Chodrichthyes | | | | | | |
| Order Charchariniformes | | | | | | |
| CHACHARINIDAE | | | | | | |
| <i>Rhizoprionodon porosus</i> (Poey, 1861) | 4 | 0.09 | 0.07 | 0.8 | 0.003 | MS |
| Order Rajiformes | | | | | | |
| NARCINIDAE | | | | | | |
| <i>Narcine brasiliensis</i> (Olfers, 1831) | 2 | 0.04 | 0.06 | 0.8 | 0.002 | MS |
| RHINOBATIDAE | | | | | | |
| <i>Rhinobatos percellens</i> (Walbaum, 1792) | 4 | 0.09 | 0.1 | 0.8 | 0.009 | MS |
| DASYATIDAE | | | | | | |
| <i>Dasyatis guttata</i> (Bloch & Schneider, 1801) | 2 | 0.04 | 1.2 | 0.8 | 0.01 | MS |
| <i>Dasyatis americana</i> (Hildebrand & Schroeder, 1928) | 3 | 0.07 | 1.3 | 0.8 | 0.02 | MS* |
| GYMNURIDAE | | | | | | |
| <i>Gymnura micrura</i> (Bloch & Schneider, 1801) | 1 | 0.02 | 0.9 | 0.4 | 0.01 | MS* |
| MYLIOBATIDAE | | | | | | |
| <i>Rhinoptera bonasus</i> (Mitchill, 1815) | 1 | 0.02 | 0.8 | 0.4 | 0.01 | MS* |
| Class Actinopterygii | | | | | | |
| Order Anguliformes | | | | | | |
| MURAENIDAE | | | | | | |
| <i>Gymnothorax ocellatus</i> Agassiz, 1831 | 1 | 0.02 | 0.008 | 0.4 | 0.001 | MS |
| <i>Gymnothorax moringa</i> (Cuvier, 1829) | 3 | 0.07 | 1.1 | 0.8 | 0.02 | MM |
| Order Albuliformes | | | | | | |
| ALBULIDAE | | | | | | |
| <i>Albula vulpes</i> (Linnaeus, 1758) | 78 | 1.94 | 0.9 | 5.9 | 0.3 | EM |
| Order Clupeiformes | | | | | | |
| ENGRAULIDAE | | | | | | |
| <i>Anchoa af. januarina</i> Hildebrand, 1943 | 44 | 0.84 | 0.7 | 4.7 | 0.2 | ER |
| <i>Anchoa lyolepis</i> (Evermann & Marsh, 1900) | 55 | 1.12 | 0.9 | 4.3 | 0.2 | ER |
| <i>Anchoa spinifer</i> (Valenciennes, 1848) (MZUFBA 5619) | 48 | 1.19 | 0.7 | 3.5 | 0.2 | ER |
| <i>Anchoa</i> sp. | 12 | 0.30 | 0.2 | 2.3 | 0.05 | ER |
| <i>Anchovia clupeioides</i> (Swainson, 1839) | 42 | 0.55 | 0.9 | 10.1 | 0.9 | MM* |
| <i>Cetengraulis edentulus</i> (Cuvier, 1829) (MZUFBA 5715 and 5951) | 105 | 2.50 | 1.98 | 10.4 | 1.1 | EM |
| <i>Lycengraulis grossidens</i> (Agassiz, 1829) | 32 | 0.79 | 0.8 | 5.4 | 0.8 | MS |
| PRISTIGASTERIDAE | | | | | | |
| <i>Pellona harroweri</i> (Fowler, 1917) | 111 | 3.75 | 4.38 | 32.4 | 3.2 | MS |
| CLUPEIDAE | | | | | | |
| <i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903) (MZUFBA 5601) | 66 | 1.64 | 0.5 | 16.7 | 1.6 | ER |
| <i>Opisthonema oglinum</i> (Lesueur, 1818) | 109 | 2.71 | 2.6 | 34.5 | 1.5 | MM* |
| <i>Harengula clupeiola</i> (Cuvier, 1829) | 54 | 1.34 | 1.2 | 15.6 | 0.5 | MS* |
| Order Siluriformes | | | | | | |
| ARIIDAE | | | | | | |
| <i>Sciades herzbergii</i> (Bloch, 1794) | 78 | 1.94 | 9.8 | 23.5 | 2.4 | ER |
| <i>Sciades couma</i> (Valenciennes, 1840) | 13 | 0.32 | 2.3 | 4.6 | 1.4 | ER |
| Order Aulopiformes | | | | | | |
| SYNODONTIDAE | | | | | | |
| <i>Synodus foetens</i> (Linnaeus, 1766) (MZUFBA 5721) | 21 | 0.52 | 0.4 | 6.8 | 0.2 | MS |

Table 1. Continued...

| Class, Order, Family and Species | N | N (%) | W (%) | C (%) | IRI | HU |
|---|-----|-------|-------|-------|-------|-----|
| Order Batrachoidiformes | | | | | | |
| BATRACHOIDIDAE | | | | | | |
| <i>Amphichthys cryptocentrus</i> (Valenciennes, 1837) | 6 | 0.15 | 0.7 | 2.2 | 0.04 | ER |
| <i>Thalassophryne nattereri</i> Steindachner, 1876 | 15 | 0.37 | 0.6 | 4.1 | 0.07 | MM |
| OGCOEPHALIDAE | | | | | | |
| <i>Ogcocephalus vespertilio</i> (Linnaeus, 1758) | 19 | 0.47 | 0.9 | 3.5 | 0.06 | MM |
| Order Mugiliformes | | | | | | |
| MUGILIDAE | | | | | | |
| <i>Mugil curema</i> Valenciennes, 1836 | 76 | 1.89 | 2.3 | 15.7 | 1.5 | EM |
| <i>Mugil liza</i> Valenciennes, 1836 | 32 | 0.79 | 1.2 | 7.6 | 0.8 | EM |
| <i>Mugil</i> sp. | 8 | 0.20 | 0.04 | 2.3 | 0.05 | EM |
| Order Atheriniformes | | | | | | |
| ATHERINOPSIDAE | | | | | | |
| <i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825) (MZUFBA 5917, 5921, 5944 and 5952) | 167 | 4.14 | 1.9 | 39.6 | 3.4 | ER |
| Order Beloniformes | | | | | | |
| BELONIDAE | | | | | | |
| <i>Strongylura timucu</i> (Walbaum, 1792) | 5 | 0.12 | 0.14 | 0.5 | 0.003 | MS |
| <i>Strongylura marina</i> (Walbaum, 1792) | 3 | 0.07 | 0.1 | 0.5 | 0.002 | MS |
| HEMIRAMPHIDAE | | | | | | |
| <i>Hyporhamphus unifasciatus</i> (Ranzani, 1841) (MZUFBA 5593) | 17 | 0.42 | 0.05 | 0.4 | 0.01 | MS |
| Order Gasterosteiformes | | | | | | |
| SYNGNATHIDAE | | | | | | |
| <i>Syngnathus</i> sp. | 3 | 0.07 | 0.01 | 1.3 | 0.01 | MS |
| <i>Hippocampus reidi</i> Ginsburg, 1933 | 6 | 0.15 | 0.03 | 1.3 | 0.02 | ER |
| FISTULARIDAE | | | | | | |
| <i>Fistularia tabacaria</i> Linnaeus, 1758 | 18 | 0.45 | 0.09 | 3.1 | 0.9 | MS* |
| Order Scorpaeniformes | | | | | | |
| DACTYLOPTERIDAE | | | | | | |
| <i>Dactylopterus volitans</i> (Linnaeus, 1758) (MZUFBA 6271) | 33 | 0.82 | 1.1 | 5.1 | 1.2 | MM* |
| SCORPAENIDAE | | | | | | |
| <i>Scorpaena plumieri</i> Bloch, 1789 | 5 | 0.12 | 0.04 | 1.3 | 0.004 | MS |
| TRIGLIDAE | | | | | | |
| <i>Prionotus punctatus</i> (Bloch, 1793) (MZUFBA 5675, 5680, 5688, 5709, 5712, 5717, 6201, 6207 and 6223) | 23 | 0.57 | 0.9 | 3.5 | 0.8 | MM* |
| Order Perciformes | | | | | | |
| MULLIDAE | | | | | | |
| <i>Pseudupeneus maculatus</i> (Bloch, 1793) | 6 | 0.15 | 0.03 | 2.3 | 0.01 | MS |
| CENTROPOMIDAE | | | | | | |
| <i>Centropomus undecimalis</i> (Bloch, 1792) (MZUFBA 5611, 5596 and 5695) | 28 | 0.69 | 1.6 | 6.7 | 1.4 | ER |
| <i>Centropomus parallelus</i> Poey, 1860 | 15 | 0.37 | 0.9 | 3.6 | 1.3 | ER |
| SERRANIDAE | | | | | | |
| <i>Diplectrum radiale</i> (Quoy & Gaimard, 1824) (MZUFBA 5600) | 21 | 0.52 | 0.04 | 4.8 | 0.04 | MM* |
| <i>Serranus flaviventris</i> (Cuvier, 1829) (MZUFBA 6225) | 19 | 0.47 | 0.01 | 5.6 | 0.01 | MM* |
| EPINEPHILIDAE | | | | | | |
| <i>Rypticus randalli</i> Couternay, 1967 (MZUFBA 5607 and 6224) | 58 | 1.44 | 0.4 | 7.5 | 0.07 | MS* |
| OPISTOGNATHIDAE | | | | | | |
| <i>Opistognathus cuvieri</i> Valenciennes, 1836 | 3 | 0.07 | 0.01 | 1.3 | 0.01 | MS |
| CARANGIDAE | | | | | | |
| <i>Oligoplites palometa</i> (Cuvier, 1832) | 19 | 0.47 | 0.7 | 7.8 | 0.09 | MM |

Table 1. Continued...

| Class, Order, Family and Species | N | N (%) | W (%) | C (%) | IRI | HU |
|---|-----|-------|-------|-------|-------|-----|
| <i>Oligoplites saliens</i> (Bloch, 1793) | 21 | 0.52 | 0.9 | 8.9 | 0.9 | MM |
| <i>Oligoplites saurus</i> (Bloch & Schneider, 1801) | 36 | 0.89 | 1.2 | 13.4 | 1.1 | MM |
| <i>Oligoplites</i> sp. | 3 | 0.07 | 0.08 | 0.4 | 0.04 | MM |
| <i>Carangoides crysos</i> (Mitchill, 1815) | 10 | 0.25 | 0.09 | 2.3 | 0.05 | MM |
| <i>Carangoides bartholomaei</i> (MZUFBA 5589) | 15 | 0.37 | 0.1 | 4.5 | 0.06 | MM |
| <i>Caranx latus</i> Agassiz, 1831 (MZUFBA 5590 and 5606) | 78 | 1.94 | 3.45 | 37.8 | 2.6 | MM* |
| <i>Caranx hippos</i> (Linnaeus, 1766) | 6 | 0.15 | 0.7 | 2.3 | 0.09 | MS |
| <i>Chloroscombrus chrysurus</i> (Linnaeus, 1766) | 66 | 1.64 | 2.34 | 25.7 | 2.4 | MS* |
| <i>Selene setapinnis</i> (Mitchill, 1815) | 4 | 0.10 | 0.03 | 0.4 | 0.03 | MS |
| <i>Selene vomer</i> (Linnaeus, 1758) | 11 | 0.27 | 0.5 | 5.6 | 0.4 | MS |
| <i>Trachinotus falcatus</i> (Linnaeus, 1758) (MZUFBA 5594) | 7 | 0.17 | 0.3 | 4.5 | 0.3 | MS |
| LUTJANIDAE | | | | | | |
| <i>Lutjanus jocu</i> (Bloch & Schneider, 1801) | 12 | 0.30 | 0.7 | 5.6 | 0.4 | MM |
| <i>Lutjanus alexandrei</i> (Moura & Lindeman, 2007) | 7 | 0.17 | 0.5 | 3.5 | 0.05 | MM |
| <i>Lutjanus synagris</i> (Linnaeus, 1758) (MZUFBA 5602 and 5942) | 43 | 1.07 | 1.2 | 19.6 | 0.9 | MM* |
| LOBOTIDAE | | | | | | |
| <i>Lobotes surinamensis</i> (Bloch, 1790) | 7 | 0.17 | 0.6 | 5.9 | 0.02 | MM |
| GERREIDAE | | | | | | |
| <i>Diapterus rhombeus</i> (Cuvier, 1829) | 117 | 2.90 | 1.8 | 28.5 | 2.5 | ER |
| <i>Diapterus auratus</i> Ranzani, 1842 | 38 | 0.94 | 0.4 | 25.7 | 0.6 | ER |
| <i>Eucinostomus gula</i> (Quoy & Gaimard, 1824) (MZUFBA 5595, 5605, 5671, 5702, 5704 and 6218) | 46 | 1.14 | 0.5 | 26.7 | 0.9 | MM |
| <i>Eucinostomus argenteus</i> Baird & Girard, 1855 (MZUFBA 5599, 5684, 5685, 5700, 5701, 5703 and 6229) | 111 | 2.76 | 0.6 | 38.8 | 2.4 | ER |
| <i>Eucinostomus melanopterus</i> (Bleeker, 1863) (MZUFBA 5592, 5597, 5705 and 6227) | 48 | 1.19 | 1 | 20.8 | 0.09 | MM* |
| <i>Eugerres brasiliensis</i> Cuvier, 1830 (MZUFBA 5614) | 16 | 0.40 | 0.07 | 3.4 | 0.06 | MM |
| <i>Gerres cinereus</i> (Walbaum, 1792) | 11 | 0.27 | 0.04 | 2.3 | 0.005 | MM* |
| HAEMULIDAE | | | | | | |
| <i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882) (MZUFBA 5608 and 5722) | 59 | 1.40 | 0.8 | 4.5 | 0.09 | MM |
| <i>Haemulon squamipinna</i> Rocha & Rosa, 1999 | 7 | 0.17 | 0.03 | 1.3 | 0.002 | MS |
| <i>Haemulon parra</i> (Desmarest, 1823) | 9 | 0.22 | 0.05 | 1.3 | 0.003 | MS |
| <i>Pomadasys corvinaeformis</i> (Steindachner, 1868) | 7 | 0.17 | 0.007 | 1.3 | 0.002 | MS |
| <i>Genyatremus luteus</i> (Bloch, 1795) (MZUFBA 5716) | 19 | 0.47 | 0.009 | 2.3 | 0.01 | ER |
| <i>Anisotremus virginicus</i> (Linnaeus, 1758) | 2 | 0.05 | 0.002 | 0.4 | 0.001 | MM |
| <i>Anisotremus surinamensis</i> | 7 | 0.17 | 0.003 | 1.3 | 0.002 | MM* |
| SPARIDAE | | | | | | |
| <i>Archosargus rhomboidalis</i> (Linnaeus, 1758) | 9 | 0.22 | 0.007 | 1.3 | 0.002 | MM |
| POLYNEMIDAE | | | | | | |
| <i>Polydactylus virginicus</i> (Linnaeus, 1758) | 67 | 1.11 | 2.11 | 33.7 | 2.3 | MS |
| SCIAENIDAE | | | | | | |
| <i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1883) | 3 | 0.07 | 0.04 | 0.8 | 0.001 | EM |
| <i>Cynoscion microlepidotus</i> (Cuvier, 1830) | 122 | 3.03 | 6.77 | 44.7 | 5.6 | ER |
| <i>Cynoscion acoupa</i> (Lacepède, 1801) | 23 | 0.57 | 0.95 | 5.6 | 0.02 | ER |
| <i>Cynoscion leiarchus</i> (Cuvier, 1830) | 74 | 1.84 | 0.9 | 10.9 | 2.9 | EM |
| <i>Isopisthus parvipinnis</i> (Cuvier, 1830) | 33 | 0.77 | 4.11 | 27.8 | 2.4 | MM* |
| <i>Larimus breviceps</i> Cuvier, 1830 | 2 | 0.05 | 0.002 | 0.4 | 0.001 | EM |
| <i>Macrodon ancylodon</i> (Bloch & Schneider, 1801) | 34 | 0.75 | 3.98 | 29.9 | 3.6 | MM* |
| <i>Menticirrhus americanus</i> (Linnaeus, 1758) | 5 | 0.12 | 0.05 | 1.3 | 0.003 | ER |
| <i>Micropogonias furnieri</i> (Desmarest, 1823) | 34 | 0.84 | 0.9 | 2.1 | 0.03 | EM |
| <i>Stellifer</i> sp. | 3 | 0.07 | 0.009 | 0.8 | 0.003 | ER |

Table 1. Continued...

| Class, Order, Family and Species | N | N (%) | W (%) | C (%) | IRI | HU |
|--|-----|-------|-------|-------|-------|-----|
| <i>Stellifer rastrifer</i> (Jordan, 1889) (MZUFBA 5718) | 178 | 4.42 | 7.89 | 59.9 | 15.5 | ER |
| <i>Stellifer stellifer</i> (Bloch, 1790) | 99 | 2.46 | 0.9 | 9.9 | 0.06 | ER |
| GOBIIDAE | | | | | | |
| <i>Bathygobius soporator</i> (Valenciennes, 1837) (MZUFBA 5615) | 49 | 1.22 | 0.21 | 5.6 | 0.05 | ER |
| <i>Gobionellus oceanicus</i> (Pallas, 1770) | 43 | 1.07 | 0.2 | 5.1 | 0.04 | ER |
| <i>Ctenogobius stomatus</i> Starks, 1913 | 135 | 3.35 | 1.04 | 50.4 | 11.7 | ER |
| <i>Ctenogobius boleosoma</i> (Jordan & Gilbert, 1882) | 57 | 1.41 | 0.2 | 5.6 | 0.06 | ER |
| <i>Ctenogobius stigmaticus</i> (Poey, 1860) | 158 | 3.92 | 1.08 | 55.7 | 10.1 | ER |
| <i>Ctenogobius smaragdus</i> (Valenciennes, 1837) | 45 | 1.12 | 0.15 | 12.3 | 0.05 | ER |
| <i>Microgobius meeki</i> Evermann & Marsh, 1900 | 4 | 0.10 | 0.001 | 1.3 | 0.002 | ER |
| EPHIPPIDAE | | | | | | |
| <i>Chaetodipterus faber</i> (Broussonet, 1782) (MZUFBA 5603) | 28 | 0.69 | 0.1 | | 0.009 | MM |
| ACANTHURIDAE | | | | | | |
| <i>Acanthurus bahianus</i> Castelnau, 1855 | 2 | 0.05 | 0.004 | 0.4 | 0.001 | MS |
| SPHYRAENIDAE | | | | | | |
| <i>Sphyraena barracuda</i> (Edwards, 1771) | 9 | 0.22 | 0.09 | 1.3 | 0.004 | MM |
| <i>Sphyraena guachancho</i> Cuvier, 1829 | 3 | 0.07 | 0.04 | 0.4 | 0.001 | MM |
| TRICHIURIDAE | | | | | | |
| <i>Trichiurus lepturus</i> Linnaeus, 1758 | 32 | 0.79 | 0.1 | 2.1 | 0.05 | EM |
| SCOMBRIDAE | | | | | | |
| <i>Scomberomorus brasiliensis</i> Collete, Russo & Zavala-Camin, 1978 | 3 | 0.07 | 0.003 | 0.8 | 0.002 | MS |
| Order Pleuronectiformes | | | | | | |
| PARALICHTHYDAE | | | | | | |
| <i>Paralichthys brasiliensis</i> (Ranzani, 1842) | 8 | 0.20 | 0.04 | 1.8 | 0.007 | MM |
| <i>Paralichthys tropicus</i> Ginsburg, 1933 | 2 | 0.05 | 0.001 | 0.4 | 0.001 | MM |
| <i>Paralichthys</i> sp. | 2 | 0.05 | 0.001 | 0.4 | 0.001 | MM |
| <i>Citharichthys spilopterus</i> Günther, 1862 | 65 | 1.61 | 0.15 | 3.4 | 0.08 | ER |
| <i>Etropus crossotus</i> Jordan & Gilbert, 1882 (MZUFBA 5609 and 5616) | 44 | 1.09 | 0.13 | 2.5 | 0.06 | ER |
| <i>Syacium micrurum</i> Ranzani, 1842 | 2 | 0.05 | 0.002 | 0.4 | 0.001 | MM |
| ACHIRIDAE | | | | | | |
| <i>Achirus lineatus</i> (Linnaeus, 1758) (MZUFBA 5620) | 31 | 0.77 | 0.15 | 13.4 | 0.1 | ER |
| <i>Achirus declives</i> Chabanaud, 1940 | 12 | 0.30 | 0.09 | 2.4 | 0.01 | ER |
| <i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915) | 54 | 1.34 | 0.14 | 22.9 | 0.3 | ER |
| <i>Trinectes microphthalmus</i> (Chabanaud, 1928) | 4 | 0.10 | 0.01 | 0.8 | 0.003 | MM |
| CYNOGLOSSIDAE | | | | | | |
| <i>Symphurus plagusia</i> (Bloch & Schneider, 1801) | 13 | 0.32 | 0.06 | 4.5 | 0.008 | MM |
| <i>Symphurus diomedianus</i> (Goode & Bean, 1885) | 2 | 0.05 | 0.001 | 0.8 | 0.001 | MS |
| Order Tetraodontiformes | | | | | | |
| MONOCANTHIDAE | | | | | | |
| <i>Stephanolepis setifer</i> (Bennett, 1831) (MZUFBA 5612) | 7 | 0.17 | 0.009 | 1.3 | 0.005 | MS |
| <i>Aluterus heudelotii</i> Hollard, 1855 | 3 | 0.07 | 0.002 | 0.4 | 0.001 | MS |
| TETRAODONTIDAE | | | | | | |
| <i>Sphoeroides testudineus</i> (Linnaeus, 1758) (MZUFBA 5672, 5673, 5674, 5678, 5679, 5682, 5683, 5687 and 5696) | 76 | 1.89 | 0.9 | 23.5 | 0.09 | ER |
| <i>Sphoeroides greeleyi</i> Gilbert, 1900 (MZUFBA 5916, 5618, 5676, 5677, 5681, 5686 and 5706) | 118 | 2.80 | 1.4 | 49.8 | 1.5 | MM* |
| <i>Sphoeroides spengleri</i> (Bloch, 1785) | 17 | 0.42 | 0.05 | 5.8 | 0.01 | MM |
| <i>Achantostracium quadricornis</i> (Linnaeus, 1758) | 7 | 0.17 | 0.001 | 4.5 | 0.006 | MS |
| DIODONTIDAE | | | | | | |
| <i>Chilomycterus spinosus</i> (Linnaeus, 1758) | 59 | 1.23 | 0.09 | 38.6 | 0.05 | MS |

The importance of the Perciformes in the present study is consistent with the pattern found in northeastern Brazil estuaries (Teixeira & Falcão 1992, Alves & Soares Filho 1996, Santos 2000, Araújo et al. 2000, Paiva et al. 2008), Bahia (Oliveira-Silva et al. 2008, Reis-Filho et al. 2010) and with inventories of the coastal zone in Southern Bahia (Moraes et al. 2009). The large number of rare species and low abundant species found in this study is consistent with the diversity pattern in a tropical environment (Pauly & Longhurst 2007). This theory can be applied in tropical estuaries where salinity fluctuations and related phenomena would also be responsible for the abundance and distribution of fish species (Vieira & Music 1993). Tropical regions are less affected by climate changing, thus there is bigger specialization of various species, favoring those with high probability of extinction to persist, resulting in a large number of rare species in the community (Giller 1984).

Evaluating the shallow areas with high prevalence of muddy sediment, there was numerical dominance of Gobiidae, Gerreidae and Tetraodontidae, however it does not fully corroborate (due to Gobiidae) with the estuaries in Southern Brazil (Corrêa 2000, Ramos & Vieira 2001, Schwarz Jr. 2005, Gomes 2005), estuary from northeast using seine net (Paiva et al. 2008) and from north utilizing block nets in mangrove tidal creek (Barletta et al. 2003). On sandy beaches in the Todos os Santos Bay near the Paraguaçu River estuary, Oliveira-Silva et al. (2008) also using seine net it was not found Gobiidae and Tetraodontidae with high abundance. Yet, Reis-Filho et al. (2010), using the same method from the authors above found Gobiidae among the four most abundant families in semi-urban estuary on the north coast of Bahia, configuring the study of estuarine fish closer to the Paraguaçu River estuary (about 220 km). But due to the acute environmental degradation (e.g. urbanization, removal of vegetation, pollution) that this system presents, comparisons of its ichthyofauna with least impacted estuaries should be made carefully.

The bottom nets accessed the ichthyofauna from the main channel dominated by Ariidae and Sciaenidae around all the Paraguaçu channel (lower estuary), similar pattern to Caeté River estuary,

Northern Brazil (Barletta et al. 2005), Sepetiba Bay, South-East Brazil (Azevedo et al. 2002) and Paranaguá estuary, Southern Brazil (Barletta et al. 2008). Although, in the middle estuary, using bottom nets, the dominant family was Sciaenidae with *Stelifer rastrifer* and *Cynoscion microlepidotus* showing higher catches and greater constancy of occurrence. Barletta et al. (2005) found *S. rastrifer* in the three sectors from Caeté River estuary (lower, middle and upper). However, in the present study this species occurred only in the middle and upper estuary preferring less saline waters. Still evaluating the shallow regions, gerreids appear as abundant and constant in the three sectors from the Formoso River estuary, Pernambuco (Paiva et al. 2008). In these last study and in estuary of the south (Falcão et al. 2006), clupeiform were abundantly collected, this group was among the most abundant on beaches near the study area (Oliveira Silva et al. 2008). In the present study, more clupeiform were caught with surface and bottom gill (5-10 m) in the middle estuary. Indeed, the dominance of gobies in shallow areas is a peculiar pattern in the structure of fish from the Paraguaçu River estuary.

There is a significant contribution of marine species in the inner estuary, most notably in mangrove creeks (Castro 2001) during a certain stage of the life cycle where the juveniles use these environments (Barletta et al. 2003, Oliveira-Neto et al. 2008). However, was observed an advancement of marine species, especially the adults (marine stragglers, marine migrants) for inner regions toward the middle estuary, showing greatest abundance and constancy of occurrence in this sector (e.g. *Caranx latus* Agassiz, 1831, *Chloroscombrus chrysurus* (Linnaeus, 1766), *Polydactylus virginicus* (Linnaeus, 1758), *Pellona harroweri*, *Harengula clupeola* (Cuvier, 1829), *Opisthonema oglinum* (Lesueur, 1818) and adults of *Macrodon ancylodon* (Bloch & Schneider, 1801) and *Isopistus parvipinis* (Cuvier, 1830)). It's accepted that this movement toward middle estuary may be due to the increased salinity in this region, and in turn the estuary did not show the three zones (upper, middle and lower) with salinity gradient set for estuaries of Brazil as shown by Barletta et al. (2005, 2008) evaluating these saline conditions. The

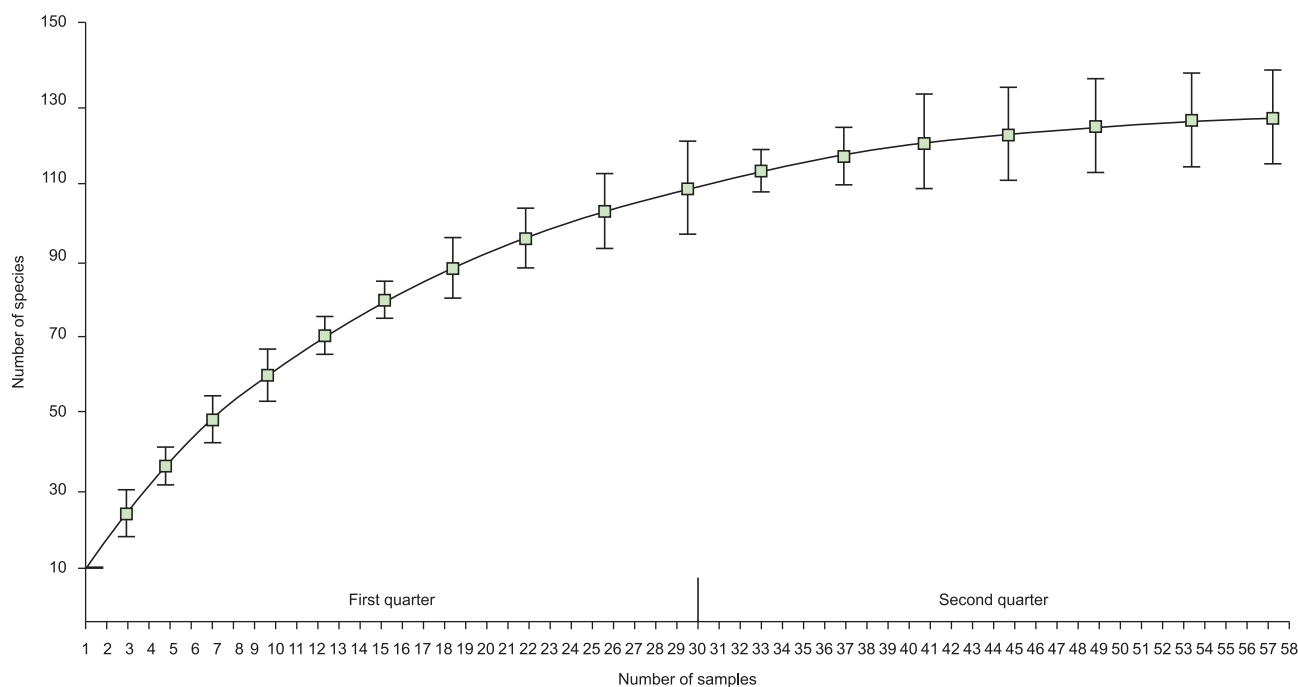


Figure 3. Species accumulation curve of the Paraguaçu River estuary by technique of rarefaction for ½ of the samples from the first and second quarter.

Figura 3. Curva de acumulação de espécies para o estuário do Rio Paraguaçu pela técnica de rarefação para ½ das amostras do primeiro e segundo trimestres.

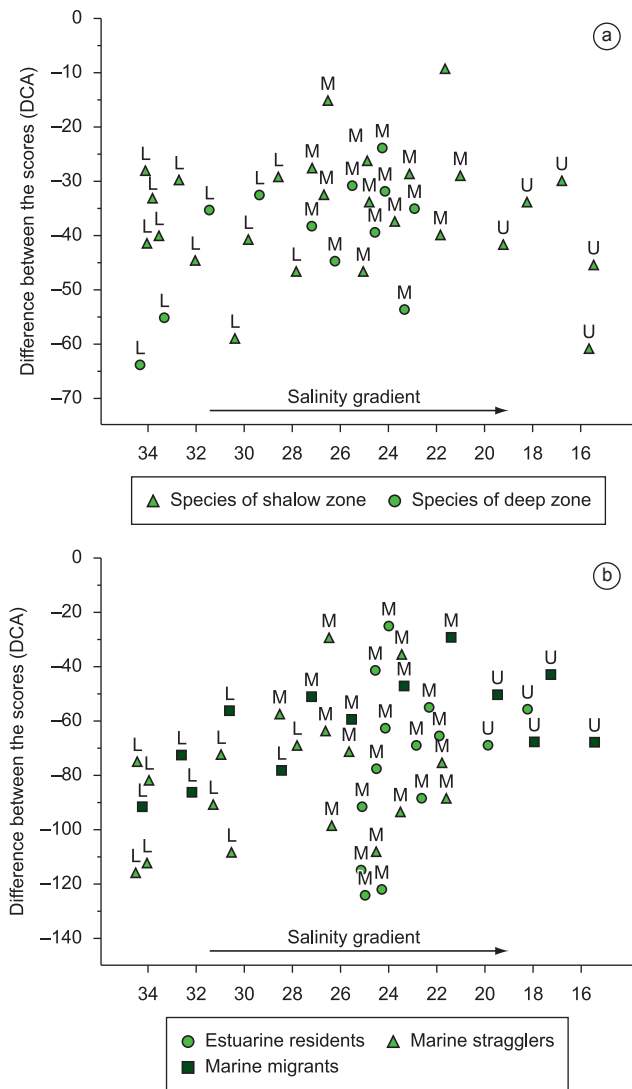


Figure 4. Scores of species derived from DCA obtained for the Paraguaçu River estuary. A: For depth zones. B: For movement of species in the estuary. (L = lower estuary, M = middle estuary; U = upper estuary).

Figura 4. Escores das espécies derivadas da DCA obtida para o estuário do Rio Paraguaçu. A: Para zonas de profundidades. B: Para o movimento das espécies no estuário. (L = baixo estuário, M = meio estuário; U = alto estuário).

ichthyofauna showed changes in distribution and structure that may be consequence of the hydrological regime imposed by the Pedra do Cavalo Dam. However, characteristics of this estuary fish community seem to be peculiar to that system as dominance of gobies in the shallow areas and Sciaenidae in middle estuary. We supported here a joint assessment of factors that influence the distribution of species within the estuary like physical-chemical gradients (Maes et al. 1998) and depth (Hyndes et al. 1999) besides those already studied as tolerance to salinity (Marshall & Elliot 1998, Barletta et al. 2005), temperature (Peterson & Ross 1991, Jaureguizar et al. 2003, Jaureguizar et al. 2004); and turbidity (Cyrus & Blaber 1992) should be incorporated into studies aiming to understand the ichthyofaunistic patterns of the Brazilian estuaries.

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